CELL
APPLICATIONS, INC.

Cytofect™-Endothelial Cell Transfection Instructions

The Cytofect™-Endothelial Cell Transfection Kit (Cat. No. TF101K) is a plasmid DNA delivery system specifically optimized to deliver DNA into a wide variety of endothelial cells. This kit contains Cytofect-2, Enhancer and Media for transfecting and culturing cells. The reagents in this kit are sufficient to perform 1000 reactions in 96-well format, 250 reactions in 24-well format, and 125 reactions in 12-well format. Peptide Enhancer is an endosomolytic peptide that complexes with DNA and Cytofect-2, then escorts the transfection complex to the nucleus. The results are high transfection efficiency (40-85%) and viability (80-95%) for those traditionally hard-to-transfect primary endothelial cells.

I. STORAGE

- ***Open the package immediately upon arrival and examine each component for shipping damage. Notify Cell Applications, Inc. or your distributor immediately if there is any problem.***

Store the transfection reagents at the following temperatures immediately upon arrival. The reagents are stable for 1 year.

Cat #	Reagent	Volume	Temp
TF52	Cytofect-2 (CF2)	250 µl	4ºC
TF53	Peptide Enhancer (PE)	750 µl	4ºC
211A	Antibiotics-Free Growth Medium	2 x 125 ml	4ºC
TF56	Transfection Medium	50 ml	4ºC

Mix each reagent well prior to use.

Cat #	Reagent	Mix
TF52	Cytofect-2 (CF2)	Invert 10 times
TF53	Peptide Enhancer (PE)	Invert 10 times

This Cytofect™-Endothelial Cell Transfection Kit contains all the necessary reagents and media for transfection of the following endothelial cells:

HAOEC (Human Aortic Endothelial Cells 304-05a)
HCAEC (Human Coronary Artery Endothelial Cells 300-05a)
HPAEC (Human Pulmonary Artery Endothelial Cells 302-05a)
HUVEC (Human Umbilical Vein Endothelial Cells 200-05n)
HLMVEC (Human Lung Microvascular Endothelial Cells 540-05a)
BAOEC (Bovine Aortic Endothelial Cells B304-05)
BCAEC (Bovine Coronary Artery Endothelial Cells G300-05)
BPAEC (Bovine Pulmonary Artery Endothelial Cells B302-05)
RAOEC (Rat Aotic Endothelial Cells R304-05)
PPAEC (Porcine Pulmonary Artery Endothelial Cells P302-05)
RBMVEC (Rat Brain Microvascular Endothelial Cells R840-05)

II. PREPARATION OF ENDOTHELIAL CELLS FOR TRANSFECTION

- ***Do not use freshly thawed cells for transfection. Cells must be passaged at least once prior to transfection.***
- ***Do not use cells that have been passaged more than 3 times prior to transfection.***
- ***Cells that have grown too crowded or sparse will yield poor results. Cell density at 75% confluence will yield the best transfection efficiency.***

1. Plate endothelial cells at a density of 25,000-30,000 cells per cm^2 in the Antibiotics-Free Growth Medium.
2. Allow cells to grow overnight. Transfect cells when cell density reaches 70-80% confluence the next day.

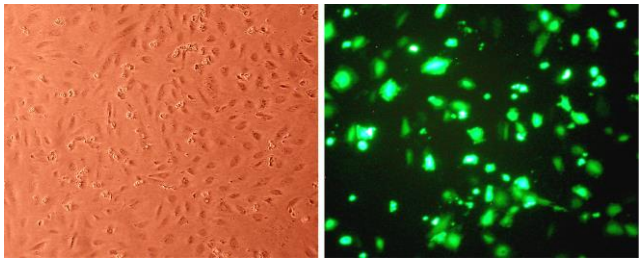

Figure 1: Transfection of HUVEC (Cat. No. 200-05) with a GFP-expression plasmid using CF2 with PE.

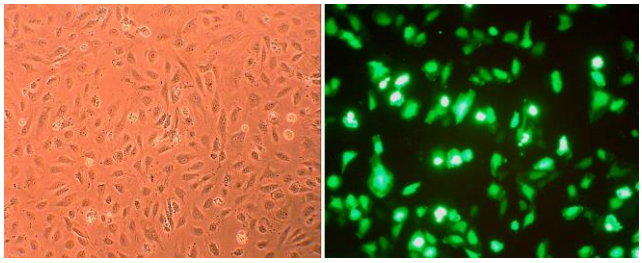

Figure 2: Transfection of HCAEC (Cat. No. 300-05) with a GFP-expression plasmid using CF2 with PE.

III. FORMATION OF TRANSFECTION COMPLEX (Table 1)

- ***Use high purity endotoxin-free DNA for transfection.***
- ***When purifying DNA, do not overload DNA purification columns with overgrown bacterial culture preparations; consult your DNA purification instructions.***
- ***Use a positive control to confirm transfection efficiency and determine the best time to carry out experiments on the transfected cells.***

A. Prepare DNA (Step 1)

1. Dilute the plasmid DNA with Transfection Medium.
2. Mix thoroughly by flicking the tube 10 times.

B. Prepare Transfection Complex (Step 2)

1. Mix CF2 thoroughly by inverting the tube 10 times.
2. Add CF2 to the diluted DNA.
3. Mix thoroughly by gently flicking 10 times.
4. Mix PE by inverting the tube 10 times.
5. Add PE to the CF2-DNA mixture.
6. Mix Transfection Cocktail thoroughly by gently flicking 10 times.
7. Incubate the Transfection Cocktail at 37°C for 25 minutes to form the Transfection Complex.

IV. TRANSFECTION OF ENDOTHELIAL CELLS (Table 2)

- ***Do not let cells dry up in the well, work only on a few wells at a time.***
- ***Use Pre-equilibrated medium for all the medium changes.***

A. Add Transfection Complex (Step 3)

1. Gently aspirate off Antibiotics-Free Growth Medium from each well.
2. Add Transfection Complex to each well by gently pipetting the Transfection Complex along the side of the well so as not to disrupt cells.
3. Incubate cells with the Transfection Complex in 37°C, 5% CO_2 humidified incubator for 1 hour.

B. Replace Transfection Complex with Antibiotic-Free Growth Medium (Step 4)

1. Gently aspirate off the Transfection Complex from each well.
2. Gently add Antibiotics-Free Growth Medium to the transfected cells in each well.
3. Incubate the transfected cells in a 37°C, 5% CO_2 humidified incubator for 24 hours.
4. Change to Endothelial Cell Growth Medium and carry out experiment.

Table 1: Formation of CF2-PE Transfection Complex

CELL APPLICATIONS, INC.	Step 1: Preparation of DNA			Step 2: Preparation of Transfection Complex					
Tissue Culture Plate	**DNA (µg)**	**Transfection Medium (µl)**		**ADD CF2 (µl)**		**ADD PE (µl)**			**Total Transfection Complex (µl)**
96-well	0.06	60	***Gently* Flick 10X to mix**	0.25	***Gently* Flick 10X to mix**	0.75	***Gently* Flick 10X to mix**	**Incubate at 37°C for 25min**	61
24-well	0.2	200		1.0		3.0			204
12-well	0.4	400		2.0		6.0			408
6-well	1.0	1000		5.0		15.0			1020

Table 2: Transfection of Endothelial Cells with CF2-PE Transfection Complex

CELL APPLICATIONS, INC.	Step 3: Addition of Transfection Complex			Step 4: Replacement of Transfection Complex with Antibiotic-Free Growth Medium		
Tissue Culture Plate		**ADD Transfection Complex (µl)**			**ADD Antibiotic-Free Growth Medium (µl)**	
96-well	**Aspirate off Antibiotic-Free Growth Medium**	61	**Incubate at 37°C 5% CO_2 for 1 hr**	**Aspirate off Transfection Complex**	100	**Incubate at 37°C 5% CO_2 for 24 hrs**
24-well		204			500	
12-well		408			1000	
6-well		1020			2000	